

Effect of Indole 3-Acetic Acid on *in vitro* germination of *Albizia amara* (Roxb.)B.

Dr.A.G.Rajalakshmi &D.Janaranjani
Assistant professor Department of Biotechnology and Research Scholar
Shri Nehru MahaVidyala College of Arts and Science,
Malumachampatti, Coimbatore-641050.

ABSTRACT

Albizia amara drought tolerant tree found in dry forests of India, belonging to the family Fabaceae. Plant growth regulators play an important role in seed germination. Auxin mediates the development processes in plants. The present study was conducted in an attempt to conserve by *in vitro* seed germination of *Albizia amara* with various concentrations of growth regulator IAA. A significant result was obtained for IAA at (5 ppm, 4 ppm and 3 ppm) compared to other treatments. Regenerated plants were successfully acclimatized and transferred to the green house with significant survival rate.

Key words: *Albizia amara*, *In vitro* seed germination, Indole-3-acetic acid.

INTRODUCTION

Albizia amara an deciduous tree, belongs to Leguminosae Gamble (1935) regeneration rate of leguminous trees in natural habitats is low. Seed germination a primary spot in plant life cycle Johnson (2000). *In vitro* tissue culture obtains a large numbers of individual Hung CD and Trueman (2010); Silva et al., (2010, 2011) medicinal plants economically important Malik et al., (1993); Prakash et al., (1994); Bhuyan et al., (1997); Arya et al., (1999); Das et al., (1999); Hussain et al., (2008) for conservation and micropropagation of *A. amara* Indravathi and Pullaiah (2013).

The leaves of *Albizia amara* contain 4'-O-menthylrutin and Budmunchiamines, sperminemacrocyclic alkaloid Mar et al., (1991) from the seeds of *Albizia amara* possess pharmaceutical activity viz., stomach cancer, common cold, diarrhoea, intestinal ailments, dandruff, wounds, skin diseases, gonorrhoea and animal fodder Ayyanar and Ignacimuthu, (2005); Kareruet al., (2008). Fruits and seed are used as anti-emetic and for treating

coughs and malaria Woongchon *et al.*, (1991). The plant extracts of *Albizia amara* are in traditional medicine Reddy *et al.*, (1967) for treating boils, eruptions and swellings, also regarded as an emetic and as a therapy for coughs, ulcer, dandruff and malaria Zhang *et al.*, (2011); Higuchi *et al.*, (1992).

Plant growth regulators enhance the growth of plants Calvo *et al.*, (2014) controlled by various environmental conditions Du Jardin (2015); Radkowski and Radkowska (2013); Colla (2014). Application of tissue culture techniques to genetic upgrading of economically important plants Scowcraft (1977). Studies on *in vitro* culture of different species of *Albizia* have been reported including *A. julibrissin* Sankhla *et al.*, (1993); *A. lebbeck* Varghese and Kaur (1988), Tomar and Gupta (1988b), *A. procera* Ghosh and Chatterjee (1992), *A. richardiana* Tomar and Gupta (1988a). *In vitro* regeneration of this species using seeds paves way for the genetic improvement of this woody species. The objectives of the present study was to evaluate the effect of seed germination of *A. amara* by *in vitro* method.

MATERIAL AND METHODS

Plant material, Media preparation and culture conditions

Seeds of *Albizia amara* were collected from Erode district, Tamil Nadu, India. The seeds were washed with distilled water for 5 times, followed by treatment of 5% sodium hypochloride for 15 min and disinfected with 0.1 % HgCl_2 for 2 min and it was rinsed four times with sterile distilled water. Experimental seeds were inoculated aseptically over Murashige and Skoog (MS) basal medium with various concentrations of IAA (0, 1, 2, 3, 4, 5 ppm) for seed germination. The cultures were maintained in the culture room at $25 \pm 2^\circ\text{C}$ under cool white fluorescent light intensity of 3000 lux.

RESULT AND DISCUSSION

The present study propagated seed in MS medium at different concentration of Indole 3-Acetic Acid. Among the five different ppm of IAA we observed the significant regeneration rate at 5 ppm, 4 ppm and 3 ppm. The plant hormone auxin has been prominently associated with numerous aspects of plant development Aloni *et al.*, (2006); Overvoorde *et al.*, (2010); Locascio *et al.*, (2014); Pattison *et al.*, (2014); Smit and Weijers (2015). Lowest germination was observed in

control. The significant seed germination was observed at 5 ppm of shoot length (6.4 cm) and root length (3.5 cm) followed by 4 ppm shoot length was observed (4.2 cm) and root length (2.3 cm). This present study supported by Kumar et al. (1998) in *Albizia procera*, Sinha, et al. (2000) in *Albizia chinensis* and Ramamurthy and Savithramma, (2003) in *Albizia amara*. Acclimatization and hardening of developed roots were thoroughly washed under tap water to remove the traces of agar from the roots and transferred to cups containing sterilized soil and sand mixture (1:1) after two weeks, the plants were transferred to larger pots containing sand and garden soil in 1:3 ratio, and were maintained under field conditions. The present study concluded that maximum germination percentage and seedling growth in *A. amara* with 5 ppm IAA.

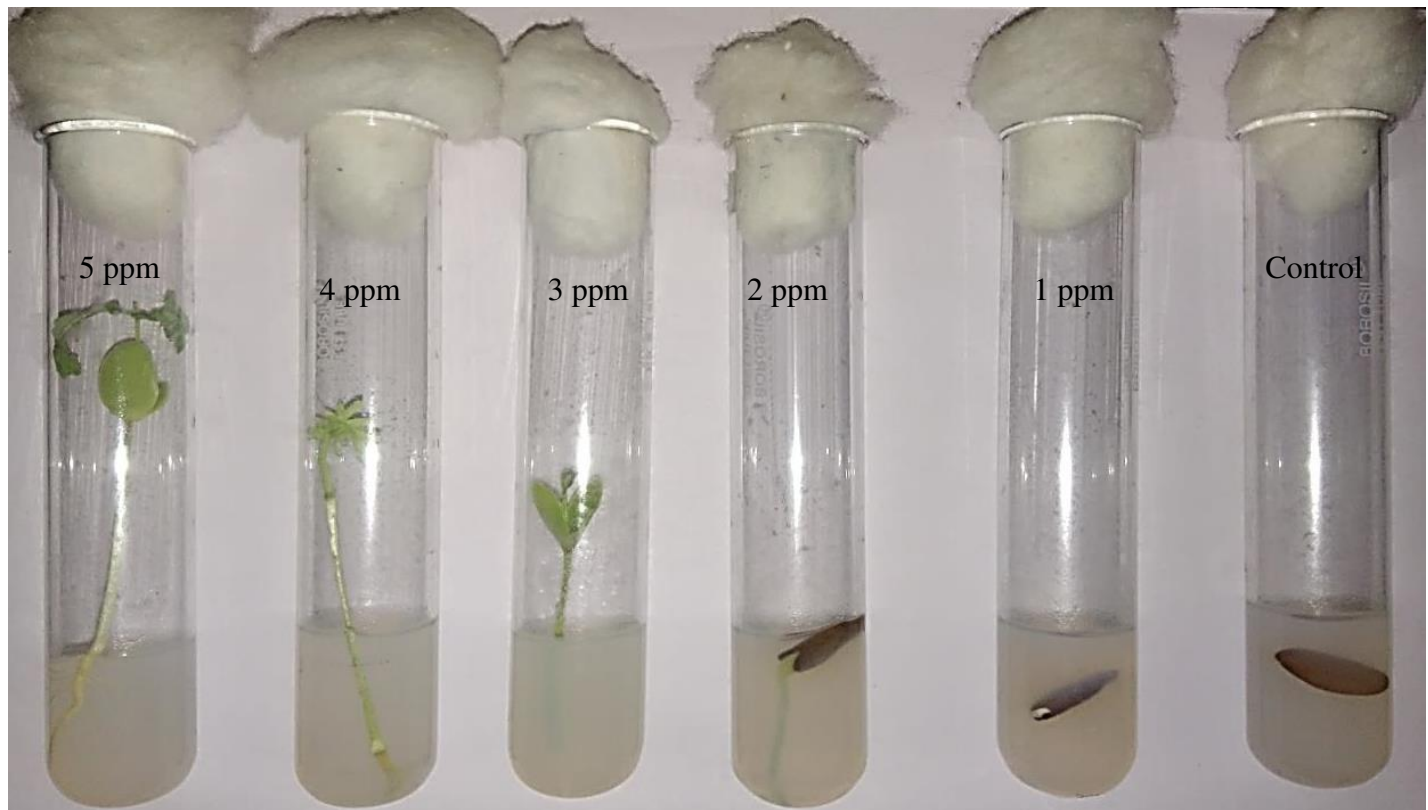




Fig.1. Effect of different concentrations IAA on of *A. amara*.

ACKNOWLEDGEMENT

The authors very sincerely acknowledge the facilities provided by Management of Shri Nehru Maha Vidyalaya College of Arts and Science, for providing facilities to do the present study.

References

- Aloni R Aloni E Langhans M Ullrich CI (2006) Role of auxin in regulating Arabidopsis flower development. *Planta*. 223: 315–328.
- Arya S Sharma S Kaur R Dev A (1999). Micropropagation of *Dendrocalamus asper* by shoot proliferation using seeds. *Plant Cell Rep*. 19: 879-882.
- Ayyanar M and Ignancimuthu S (2005) Medicinal Plants used by the tribals of Tirunelveli hills, Tamilnadu to treat poisonous bites and skin diseases. *Indian J of Traditional Knowledge*. 4: 229-236.
- Bhuyan KK Pattanaik S Chand PK (1997) Micropropagation of Curry leaf tree [*Murraya koenigii* (L.) Spreng.] by axillary proliferation using intact seedlings. *Plant Cell Rep*. 16: 779-782.
- Calvo P Nelson L Kloepper JW (2014) Agricultural uses of plant biostimulants. *Plant Soil*. 383: 3–41.
- Colla G Roupael Y Canaguier R Svecova E Cardarelli M (2014) Biostimulant action of a plant-derived protein hydrolysate produced through enzymatic hydrolysis. *Front. Plant Sci*. 5: 448.
- Das DK Prakash NS Bhalla-Sarin N (1999). Multiple shoot induction and plant regeneration in litchi (*Litchi chinensis* Sonn.). *Plant Cell Rep*. 18: 691-695.
- Du Jardin P (2015) Plant biostimulants: Definition, concept, main categories and regulation. *Sci. Hort*. 196: 3–14.

Gamble JS (1935) The Flora of the Presidency of Madras, Adlard & Son, Ltd, London.

Ghosh N and Chatterjee A (1992) Cytological studies on the effect of gamma radiation on *Albizia procera* Benth. In: G.K. Manna and S.C. Roy (eds.). Perspectives in Cytology and Genetics. 7: 1199-1209.

Ghosh N and Chatterjee A (1992) Cytological study on the effect of aging on callus cultures of *Dalbergia sissoo* Roxb. Plant Sciences in the Nineties. 440-447.

Higuchi H Kinjo J Nohara T (1992) An arrhythmic-inducing glycoside from *Albizia julibrissin* Durazz. IV. Chemical and Pharmaceutical Bulletin. 40: 829– 831.

Hung CD and Trueman SJ (2010) Nutrient responses differ between node and organogenic cultures of *Corymbia torelliana* x *C. citriodora* (Myrtaceae). Australian Journal of Botany. 58: 410–419.

Hussain TM, Chandrasekhar T, Gopal GR, 2008. Micropropagation of *Sterculia aurea* Roxb., an endangered tree species from intact seedlings. Afr J Biotechnol 7: 095-101.

Indravathi G and Pullaiah T (2013) In vitro propagation studies of *Albizia amara* (Roxb.) Boiv. African Journal of Plant Science. 7: 1-8.

Johnson R (2000) Characterization of germination related genes in *Avena fatua* L. seeds. In: Seed Biology Advances and Applications: Proceedings of the Sixth International Workshop on Seeds. Black M, Bradford KJ, Vazquez- Ramos J (Eds.) CABI Publishing, Mérida, México.

Kareru PG Gachanja AN Keriko JM Kenji GM (2008) Antimicrobial activity of some medicinal plants used by herbalists in eastern province. African Journal of Traditional, Complementary and Alternative Medicines. 5: 51-55.

Kumar S Sarkar AK Kunhi Kannan (1998) Regeneration of plants from leaflet explants of tissue culture raised safedsiris *Albizia procera*. Plant Cell Tissue and Organ Culture. 54: 137– 143.

Locascio A Roig-Villanova I Bernardi J Varotto S (2014) Current perspectives on the hormonal control of seed development in *Arabidopsis* and maize: a focus on auxin. Front Plant Sci. 5: 412

Malik AK Ali-Khan ST Saxena PK (1993) High-frequency organogenesis from direct seed culture of *Lathyrus*. Ann Bot. 72: 629-637.

Mar W Tan GT Cordell GA Pezzuto JM (1991). Biological activity of novel macrocyclic alkaloids (Budmunchiamines) from *Albizia amara* detected on the basis of interaction with DNA. Journal of Natural Products. 54: 1531-42.
Mental Biology – Plant. 36: 370-373.

Overvoorde P Fukaki H Beeckman T (2010). Auxin control of root development. Cold Spring Harb Perspect Biol. 2: 1537,

Pattison RJ Csukasi F Catalá C (2014) Mechanisms regulating auxin action during fruit development. *Physiol Plant*. 151: 62–72

Prakash SN Pental D Sarin NB (1994). Regeneration of pigeon pea (*Cajanus cajan*) from cotyledonary node via multiple shoot formation. *Plant Cell Rep*. 13: 623-627.

Radkowski A and Radkowska I (2013) Effect of foliar application of growth biostimulant on quality and nutritive value of meadow sward. *Ecol. Chem. Eng. A*. 20: 1205–1211.

Ramamurthy N and Savithramma N (2003). Shoot bud regeneration from leaf explants of *Albizia amara* Boiv. *Indian J. Plant Physiol*. 8: 372- 376.

Reddy Sastry CV Rukmini C Ramachandra Rao L (1967) Chemistry of Saponins : part III – Isolation of new flavonol Glycoside, 4'-O- Methylquercetin-3-rutinoside, from *Albizia amara* Benth. *Indian Journal of Chemistry*. 5.
regeneration of *Albizia chinensis* (Osbeck) Merr. *In Vitro Cellular and Develop-*

Sankhla D Davis TD Sankhla N (1993) Effect of gibberellin biosynthesis inhibitors on shoot regeneration from hypocotyl explants of *Albizia julibrissin*. *Plant Cell Report*. 13: 115-118.

Sgowcraft W (1977) Somatic cell genetics and plant improvement. *Ann. Rev. Agron*. 29: 39-81.

Silva ALL Oliveira Y Costa JL (2010) Shoot tip and cotyledon explants of *Eucalyptus saligna* Sm. cultivated on different kanamycin levels. *Journal of Biotechnology and Biodiversity*. 1: 1–5.

Silva ALL Oliveira Y Costa JL (2011) Preliminary results for genetic transformation of shoot tip of *Eucalyptus saligna* Sm. via *Agrobacterium tumefaciens*. *Journal of Biotechnology and Biodiversity*. 2: 1–6.

Sinha RK Majundar K Sinha S (2000) In vitro differentiation and plant

Smit ME and Weijers D (2015) The role of auxin signaling in early embryo pattern formation. *Curr Opin Plant Biol*. 28: 99–105.

Tomar UK and SC Gupta (1988b) In vitro plant regeneration of leguminous trees (*Albizia* sp.) *Plant Cell Rep*. 7: 385-388.

Tomar UK and SC Gupta (1988a) Somatic embryogenesis and organogenesis in callus cultures of a tree legume, *Albizia richardiana* King. *Plant Cell Rep*. 7: 70-73.

Varghese TM and Kaur A (1988) In vitro propagation of *Albizia lebbek* Benth. *Curr. Sci*. 57: 1010-1012.

Woongchon M Ghee TT Geoffrey AC John MP (1991) Biological activity of novel macrocyclic alkaloids (budmunchiamines) from *Albizia amara* detected on the basis of interaction with DNA. *Journal of Natural Products*. 54: 1531-42.

Zhang H Samadi AK Rao KV Cohen MS Timmermann BN (2011). Cytotoxic oleanane-type saponins from *Albizia inundata*. *Journal of Natural Products*. 74: 477-82.